

Envirolab Services Pty Ltd ABN 37 112 535 645

12 Ashley St Chatswood NSW 2067 ph 02 9910 6200 fax 02 9910 6201 customerservice@envirolab.com.au www.envirolab.com.au

### **CERTIFICATE OF ANALYSIS 369940**

Client Details	
Client	NSW Health
Attention	Kwendy Cavanagh
Address	Locked Bag 2030, ST LEONARDS, NSW, 1590

Sample Details	
Your Reference	Narrabri Shire Council - Killarney Bore
Number of Samples	3 Water
Date samples received	07/01/2025
Date completed instructions received	07/01/2025

### **Analysis Details**

Please refer to the following pages for results, methodology summary and quality control data.

Samples were analysed as received from the client. Results relate specifically to the samples as received.

Results are reported on a dry weight basis for solids and on an as received basis for other matrices.

Report Details	
Date results requested by	14/01/2025
Date of Issue	09/01/2025
NATA Accreditation Number 2901.	This document shall not be reproduced except in full.
Accredited for compliance with ISO	/IEC 17025 - Testing. Tests not covered by NATA are denoted with *

Results Approved By

Sean McAlary, Senior Chemist

<u>Authorised By</u> Nancy Zhang, Laboratory Manager

Envirolab Reference: 369940 Revision No: R00



Our Reference		369940-1
Your Reference	UNITS	Killamey Bore A24NA0100008
Barcode		A24NA0100008
Sample Site Code		N29
Date Sampled		6/01/2025
Type of sample		Water
Date prepared	-	08/01/2025
Date analysed		08/01/2025
Perfluorobutanesulfonic acid	μg/L	0.005
Perfluorohexanesulfonic acid - PFHxS	μg/L	0.025
Perfluorooctanesulfonic acid PFOS	µg/L	0.029
Perfluorooctanoic acid PFOA	µg/L	0.002
6:2 FTS	μg/L	<0.001
8:2 FTS	µg/L	<0.002
Surrogate 13 C <sub>8</sub> PFOS	%	104
Surrogate 13 C2 PFOA	%	87
Extracted ISTD 13 C <sub>3</sub> PFBS	%	72
Extracted ISTD 18 O2 PFHxS	%	97
Extracted ISTD 13 C <sub>4</sub> PFOS	%	87
Extracted ISTD 13 C4 PFOA	%	107
Extracted ISTD <sup>13</sup> C₂ 6:2FTS	%	111
Extracted ISTD <sup>13</sup> C <sub>2</sub> 8:2FTS	%	115
Total Positive PFHxS & PFOS	μg/L	0.053
Total Positive PFOA & PFOS	μg/L	0.031
Total Positive PFAS	μg/L	0.061

Envirolab Reference: 369940

Revision No:

Method ID	Methodology Summary
Org-029	Soil samples are extracted with basified Methanol. Waters and soil extracts are directly injected and/or concentrated/extracted using SPE. TCLPs/ASLP leachates are centrifuged, the supernatant is then analysed (including amendment with solvent) - as per the option in AS4439.3.
	Analysis is undertaken with LC-MS/MS.
	PFAS results include the sum of branched and linear isomers where applicable.
	Please note that PFAS results are corrected for Extracted Internal Standards (QSM 5.4 Table B-15 terminology), which are mass labelled analytes added prior to sample preparation to assess matrix effects and verify processing of the sample. PFAS analytes without a commercially available mass labelled analogue are corrected vs a closely eluting mass labelled PFAS compound. Surrogates are also reported, in this context they are mass labelled PFAS compounds added prior to extraction be are used as monitoring compounds only (not used for result correction). Envicarb (or similar) is used discretionally to remove interfering matrix components.
	Please contact the laboratory if estimates of Measurement Uncertainty are required as per WA DER.

Envirolab Reference: 369940 R00

Page | 3 of 6

QUALITY CONTROL: PFAS in Water LOW LEVEL Short					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date prepared	-			08/01/2025	U				07/01/2025	
Date analysed	-			08/01/2025			lyti -		07/01/2025	
Perfluorobutanesulfonic acid	μg/L	0.001	Org-029	<0.001				1 -	102	
Perfluorohexanesulfonic acid - PFHxS	μg/L	0.001	Org-029	<0.001	III,		100	15-1	101	
Perfluorooctanesulfonic acid PFOS	μg/L	0.001	Org-029	<0.001			Hat -		100	
Perfluorooctanoic acid PFOA	µg/L	0.001	Org-029	<0.001			- w-		98	
S:2 FTS	μg/L	0.001	Org-029	<0.001	171			1	98	
3:2 FTS	μg/L	0.002	Org-029	<0.002	12:31		147	· (we)	100	
Surrogate 13 C <sub>8</sub> PFOS	%		Org-029	98	-21		the fi		98	
Suπogate <sup>13</sup> C <sub>2</sub> PFOA	%		Org-029	91	lipj.		MIT	TATE !	93	
Extracted ISTD 13 C <sub>3</sub> PFBS	%		Org-029	70			INTE	IIVY	64	
Extracted ISTD 18 O2 PFHxS	%		Org-029	80	eye (		hett,	PIL	77	
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOS	%		Org-029	72	urs.		4.1	11,23	71	
extracted ISTD 13 C <sub>4</sub> PFOA	%		Org-029	89	en û		ner)	IVT	84	
extracted ISTD 13 C <sub>2</sub> 6:2FTS	%		Org-029	95	N=F	19/2	TXTE:	[[07]	83	
xtracted ISTD 13 C <sub>2</sub> 8:2FTS	%		Org-029	93			DETE	100	83	

Envirolab Reference: 369940 Revision No: R00

Result Definiti	ons
NT	Not tested
NA	Test not required
INS	Insufficient sample for this test
PQL	Practical Quantitation Limit
<	Less than
>	Greater than
RPD	Relative Percent Difference
LCS	Laboratory Control Sample
NS	Not specified
NEPM	National Environmental Protection Measure
NR	Not Reported

Envirolab Reference: **369940** Revision No: **R00** 

Quality Contro	ol Definitions
Blank	This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.
Duplicate	This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.
Matrix Spike	A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.
LCS (Laboratory Control Sample)	This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.
Surrogate Spike	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.
Australian Drinking	Mater Guidelines recommend that Thermatelerent Californ, Ferent Februaria, R. F. California, I. C.

Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011.

The recommended maximums for analytes in urine are taken from "2018 TLVs and BEIs", as published by ACGIH (where available). Limit provided for Nickel is a precautionary guideline as per Position Paper prepared by AIOH Exposure Standards Committee, 2016.

Guideline limits for Rinse Water Quality reported as per analytical requirements and specifications of AS 4187, Amdt 2 2019, Table 7.2

### Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was insufficient in order to satisfy laboratory QA/QC protocols.

When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Where matrix spike recoveries fall below the lower limit of the acceptance criteria (e.g. for non-labile or standard Organics <60%), positive result(s) in the parent sample will subsequently have a higher than typical estimated uncertainty (MU estimates supplied on request) and in these circumstances the sample result is likely biased significantly low.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

Samples for Microbiological analysis (not Amoeba forms) received outside of the 2-8°C temperature range do not meet the ideal cooling conditions as stated in AS2031-2012.

Envirolab Reference: 369940



#### **Envirolab Services Pty Ltd**

ABN 37 112 535 645 12 Ashley St Chatswood NSW 2067 ph 02 9910 6200 fax 02 9910 6201 customerservice@envirolab.com.au www.envirolab.com.au

### **CERTIFICATE OF ANALYSIS 369938**

Client Details	
Client	NSW Health
Attention	Kwendy Cavanagh
Address	Locked Bag 2030, ST LEONARDS, NSW, 1590

Sample Details	
Your Reference	Narrabri Shire Council - Tibberena Bore
Number of Samples	3 Water
Date samples received	07/01/2025
Date completed instructions received	07/01/2025

### **Analysis Details**

Please refer to the following pages for results, methodology summary and quality control data.

Samples were analysed as received from the client. Results relate specifically to the samples as received.

Results are reported on a dry weight basis for solids and on an as received basis for other matrices.

Report Details		
Date results requested by	14/01/2025	
Date of Issue	09/01/2025	
NATA Accreditation Number 2901.	This document shall not be reproduced except in full.	
	/IEC 17025 - Testing. Tests not covered by NATA are denoted with *	

Results Approved By
Sean McAlary, Senior Chemist

Authorised By

Nancy Zhang, Laboratory Manager

Envirolab Reference: 369938 Revision No: R00



Our Reference		369938-1	369938-2	369938-3
Your Reference	UNITS	Tibberena Bore A24NA0100007	Tibberena Bore Duplicate	Tibberena Bore Field Blank
Barcode		A24NA0100007		
Sample Site Code	T V NE	N28	- 1	*
Date Sampled	1 1 1	6/01/2025	6/01/2025	6/01/2025
Type of sample		Water	Water	Water
Date prepared	=	08/01/2025	08/01/2025	08/01/2025
Date analysed	4	08/01/2025	08/01/2025	08/01/2025
Perfluorobutanesulfonic acid	µg/L	0.001	0.001	<0.001
Perfluorohexanesulfonic acid - PFHxS	µg/L	0.007	0.007	<0.001
Perfluorooctanesulfonic acid PFOS	µg/L	0.011	0.011	<0.001
Perfluorooctanoic acid PFOA	μg/L	<0.001	<0.001	<0.001
5:2 FTS	μg/L	<0.001	<0.001	<0.001
3:2 FTS	μg/L	<0.002	<0.002	<0.002
Surrogate 13 C <sub>8</sub> PFOS	%	93	105	102
Surrogate 13 C <sub>2</sub> PFOA	%	89	95	94
Extracted ISTD <sup>13</sup> C <sub>3</sub> PFBS	%	70	74	75
Extracted ISTD 18 O2 PFHxS	%	93	89	92
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOS	%	92	87	86
Extracted ISTD 13 C4 PFOA	%	103	100	99
Extracted ISTD <sup>13</sup> C <sub>2</sub> 6:2FTS	%	108	113	104
Extracted ISTD 13 C2 8:2FTS	%	112	116	105
Total Positive PFHxS & PFOS	μg/L	0.018	0.018	<0.001
otal Positive PFOA & PFOS	μg/L	0.011	0.011	<0.001
Total Positive PFAS	μg/L	0.019	0.020	<0.001

Envirolab Reference: 369938 R00 Revision No:

Method 1D	Methodology Summary
Org-029	Soil samples are extracted with basified Methanol. Waters and soil extracts are directly injected and/or concentrated/extracted using SPE. TCLPs/ASLP leachates are centrifuged, the supernatant is then analysed (including amendment with solvent) - as per the option in AS4439.3.
	Analysis is undertaken with LC-MS/MS.
	PFAS results include the sum of branched and linear isomers where applicable.
	Please note that PFAS results are corrected for Extracted Internal Standards (QSM 5.4 Table B-15 terminology), which are mass labelled analytes added prior to sample preparation to assess matrix effects and verify processing of the sample. PFAS analytes without a commercially available mass labelled analogue are corrected vs a closely eluting mass labelled PFAS compound. Surrogates are also reported, in this context they are mass labelled PFAS compounds added prior to extraction but are used as monitoring compounds only (not used for result correction). Envicarb (or similar) is used discretionally to remove interfering matrix components.
	Please contact the laboratory if estimates of Measurement Uncertainty are required as per WA DER,

Enviroiab Reference; 369938 Revision No: R00

QUALITY CONTR	OL: PFAS i	n Water LO	W LEVEL Shor	t		Du	plicate		Spike Re	covery %
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	369938-1
Date prepared	-			08/01/2025	111				07/01/2025	08/01/2025
Date analysed				08/01/2025	ALI I		24.1		07/01/2025	08/01/2025
Perfluorobutanesulfonic acid	μg/L	0.001	Org-029	<0.001					102	103
Perfluorohexanesulfonic acid - PFHxS	μg/L	0.001	Org-029	<0.001			- 0		101	96
Perfluorooctanesulfonic acid PFOS	μg/L	0.001	Org-029	<0.001	ш		14		100	96
Perfluorooctanoic acid PFOA	µg/L	0.001	Org-029	<0.001				100	98	100
3:2 FTS	μg/L	0.001	Org-029	<0.001					98	99
3:2 FTS	μg/L	0.002	Org-029	<0.002	garage.		191	100	100	106
Surrogate 13 C <sub>8</sub> PFOS	%		Org-029	98	- 1				98	97
Suπogate <sup>13</sup> C₂ PFOA	%		Org-029	91	10		DI.	200	93	90
Extracted ISTD <sup>13</sup> C <sub>3</sub> PFBS	%		Org-029	70			1917		64	68
Extracted ISTD <sup>18</sup> O₂ PFHxS	%		Org-029	80			ηĦ.	L red	77	91
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOS	%		Org-029	72			14		71	87
extracted ISTD 13 C4 PFOA	%		Org-029	89	m),	TXT]	MIT	ļ pr	84	96
extracted ISTD <sup>13</sup> C <sub>2</sub> 6:2FTS	%		Org-029	95	1		1675	65	83	99
xtracted ISTD 13 C <sub>2</sub> 8:2FTS	%		Org-029	93	(MTH		9/11	De CLU	83	92

Envirolab Reference: 369938 Revision No: R00

Result Definiti	ons
NT	Not tested
NA	Test not required
INS	Insufficient sample for this test
PQL	Practical Quantitation Limit
<	Less than
>	Greater than
RPD	Relative Percent Difference
LCS	Laboratory Control Sample
NS	Not specified
NEPM	National Environmental Protection Measure
NR	Not Reported

Envirolab Reference: 369938 Revision No: R00

Quality Contro	ol Definitions
Blank	This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.
Duplicate	This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.
Matrix Spike	A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.
LCS (Laboratory Control Sample)	This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.
Surrogate Spike	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.

Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011.

The recommended maximums for analytes in urine are taken from "2018 TLVs and BEIs", as published by ACGIH (where available). Limit provided for Nickel is a precautionary guideline as per Position Paper prepared by AIOH Exposure Standards Committee, 2016.

Guideline limits for Rinse Water Quality reported as per analytical requirements and specifications of AS 4187, Amdt 2 2019, Table 7.2

### Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was insufficient in order to satisfy laboratory QA/QC protocols.

When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Where matrix spike recoveries fall below the lower limit of the acceptance criteria (e.g. for non-labile or standard Organics <60%), positive result(s) in the parent sample will subsequently have a higher than typical estimated uncertainty (MU estimates supplied on request) and in these circumstances the sample result is likely biased significantly low.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

Samples for Microbiological analysis (not Amoeba forms) received outside of the 2-8°C temperature range do not meet the ideal cooling conditions as stated in AS2031-2012.

Envirolab Reference: 369938

Revision No:



Envirolab Services Pty Ltd
ABN 37 112 535 645
12 Ashley St Chatswood NSW 2067
ph 02 9910 6200 fax 02 9910 6201
customerservice@envirolab.com.au
www.envirolab.com.au

### **CERTIFICATE OF ANALYSIS 369937**

Client Details	
Client	NSW Health
Attention	Kwendy Cavanagh
Address	Locked Bag 2030, ST LEONARDS, NSW, 1590

Sample Details	
Your Reference	Narrabri Shire Council - Elizabeth Bore
Number of Samples	3 Water
Date samples received	07/01/2025
Date completed instructions received	07/01/2025

## **Analysis Details**

Please refer to the following pages for results, methodology summary and quality control data.

Samples were analysed as received from the client. Results relate specifically to the samples as received.

Results are reported on a dry weight basis for solids and on an as received basis for other matrices.

Report Details		
Date results requested by	14/01/2025	
Date of Issue	09/01/2025	
NATA Accreditation Number 2901.	This document shall not be reproduced except in full.	
Accredited for compliance with ISO	IEC 17025 - Testing. Tests not covered by NATA are denoted with *	

Results Approved By
Sean McAlary, Senior Chemist

Authorised By
Nancy Zhang, Laboratory Manager

Envirolab Reference: 369937 Revision No: R00



Our Reference		369937-1
Your Reference	UNITS	Elizabeth Bore A24NA0100006
Barcode	-	A24NA0100006
Sample Site Code		N30
Date Sampled		6/01/2025
Type of sample		Water
Date prepared		08/01/2025
Date analysed	*	08/01/2025
Perfluorobutanesulfonic acid	µg/L	<0.001
Perfluorohexanesulfonic acid - PFHxS	μg/L	0.002
Perfluorooctanesulfonic acid PFOS	μg/L	0.006
Perfluorooctanoic acid PFOA	µg/L	<0.001
6:2 FTS	µg/L	<0.001
8:2 FTS	µg/L	<0.002
Surrogate <sup>13</sup> C <sub>8</sub> PFOS	%	105
Surrogate <sup>13</sup> C₂ PFOA	%	90
Extracted ISTD 13 C3 PFBS	%	75
Extracted ISTD 18 O2 PFHxS	%	94
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOS	%	90
Extracted ISTD 13 C4 PFOA	%	108
Extracted ISTD 13 C2 6:2FTS	%	115
Extracted ISTD 13 C2 8:2FTS	%	116
Total Positive PFHxS & PFOS	μg/L	0.008
Total Positive PFOA & PFOS	μg/L	0.006
Total Positive PFAS	μg/L	0.008

Envirolab Reference: 369937 Revision No: R00

Method ID	Methodology Summary
Org-029	Soil samples are extracted with basified Methanol. Waters and soil extracts are directly injected and/or concentrated/extracted using SPE. TCLPs/ASLP leachates are centrifuged, the supernatant is then analysed (including amendment with solvent) - as per the option in AS4439.3.
	Analysis is undertaken with LC-MS/MS.
	PFAS results include the sum of branched and linear isomers where applicable.
	Please note that PFAS results are corrected for Extracted Internal Standards (QSM 5.4 Table B-15 terminology), which are mass labelled analytes added prior to sample preparation to assess matrix effects and verify processing of the sample. PFAS analytes without a commercially available mass labelled analogue are corrected vs a closely eluting mass labelled PFAS compound. Surrogates are also reported, in this context they are mass labelled PFAS compounds added prior to extraction but are used as monitoring compounds only (not used for result correction). Envicarb (or similar) is used discretionally to remove interfering matrix components.
	Please contact the laboratory if estimates of Measurement Uncertainty are required as per WA DER.

Envirolab Reference: 369937 Revision No: R00

QUALITY CONTR	OL: PFAS i	n Water LO	W LEVEL Short			Du	plicate		Spike Rec	covery %
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date prepared	-			08/01/2025	1	08/01/2025	08/01/2025		07/01/2025	
Date analysed	-			08/01/2025	1	08/01/2025	08/01/2025		07/01/2025	
Perfluorobutanesulfonic acid	μg/L	0.001	Org-029	<0.001	1	<0.001	<0.001	0	102	
Perfluorohexanesulfonic acid - PFHxS	µg/L	0.001	Org-029	<0.001	1	0.002	0.002	0	101	
Perfluorooctanesulfonic acid PFOS	μg/L	0.001	Org-029	<0.001	1	0.006	0.005	18	100	
Perfluorooctanoic acid PFOA	μg/L	0.001	Org-029	<0.001	1	<0.001	<0.001	0	98	
9:2 FTS	μg/L	0.001	Org-029	<0.001	1	<0.001	<0.001	0	98	
3:2 FTS	µg/L	0.002	Org-029	<0.002	1	<0.002	<0,002	0	100	
Surrogate <sup>13</sup> C <sub>8</sub> PFOS	%		Org-029	98	1	105	98	7	98	
Suπogate <sup>13</sup> C₂ PFOA	%	1 -	Org-029	91	1	90	89	1	93	
Extracted ISTD <sup>13</sup> C <sub>3</sub> PFBS	%		Org-029	70	1	75	67	11	64	
Extracted ISTD <sup>18</sup> O <sub>2</sub> PFHxS	%		Org-029	80	1	94	96	2	77	
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOS	%		Org-029	72	1	90	90	0	71	
Extracted ISTD 13 C <sub>4</sub> PFOA	%		Org-029	89	1	108	107	1	84	
Extracted ISTD <sup>13</sup> C <sub>2</sub> 6:2FTS	%		Org-029	95	1	115	113	2	83	
Extracted ISTD <sup>13</sup> C <sub>2</sub> 8:2FTS	%		Org-029	93	1	116	115	1	83	

Envirolab Reference: 369937 Revision No: R00

<b>Quality Control</b>	ol Definitions
Blank	This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.
Duplicate	This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.
Matrix Spike	A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.
LCS (Laboratory Control Sample)	This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.
Surrogate Spike	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.
	the second secon

Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011

The recommended maximums for analytes in urine are taken from "2018 TLVs and BEIs", as published by ACGIH (where available). Limit provided for Nickel is a precautionary guideline as per Position Paper prepared by AIOH Exposure Standards Committee, 2016.

Guideline limits for Rinse Water Quality reported as per analytical requirements and specifications of AS 4187, Amdt 2 2019, Table 7 2

### Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was insufficient in order to satisfy laboratory QA/QC protocols.

When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Where matrix spike recoveries fall below the lower limit of the acceptance criteria (e.g. for non-labile or standard Organics <60%), positive result(s) in the parent sample will subsequently have a higher than typical estimated uncertainty (MU estimates supplied on request) and in these circumstances the sample result is likely biased significantly low.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

Samples for Microbiological analysis (not Amoeba forms) received outside of the 2-8°C temperature range do not meet the ideal cooling conditions as stated in AS2031-2012.

Envirolab Reference: 369937 Revision No: R00

NT	Not tested
NA	Test not required
INS	Insufficient sample for this test
PQL	Practical Quantitation Limit
<	Less than
>	Greater than
RPD	Relative Percent Difference
LCS	Laboratory Control Sample
NS	Not specified
NEPM	National Environmental Protection Measure
NR	Not Reported

Envirolab Reference: 369937 Revision No R00



#### **Envirolab Services Pty Ltd**

ABN 37 112 535 645
12 Ashley St Chatswood NSW 2067
ph 02 9910 6200 fax 02 9910 6201
customerservice@envirolab.com.au
www.envirolab.com.au

### **CERTIFICATE OF ANALYSIS 369941**

Client Details	
Client	NSW Health
Attention	Kwendy Cavanagh
Address	Locked Bag 2030, ST LEONARDS, NSW, 1590

Sample Details						
Your Reference	Narrabri Shire Council - Namoi Reservoir					
Number of Samples	3 Water					
Date samples received	07/01/2025					
Date completed instructions received	07/01/2025					

### **Analysis Details**

Please refer to the following pages for results, methodology summary and quality control data.

Samples were analysed as received from the client. Results relate specifically to the samples as received.

Results are reported on a dry weight basis for solids and on an as received basis for other matrices.

/2025			,																
/2025			;																
nt shall not b	ot i	not	ıll not	not b	t be i	е гер	repro	produ	luced	exc	cept	t in f	ull.						
nt shall not b Testing. <b>Tes</b>														don	otod :	with	*		

Results Approved By

Sean McAlary, Senior Chemist

**Authorised By** 

Nancy Zhang, Laboratory Manager

Envirolab Reference: 369941 Revision No. R00



Our Reference		369941-1
Your Reference	UNITS	Namoi Reservoid A24NA0100009
Barcode		A24NA0100009
Sample Site Code	Mary Mary	N27
Date Sampled		8/01/2025
Type of sample		Water
Date prepared	-	08/01/2025
Date analysed	-	08/01/2025
Perfluorobutanesulfonic acid	µg/L	0.001
Perfluorohexanesulfonic acid - PFHxS	μg/L	0.007
Perfluorooctanesulfonic acid PFOS	μg/L	0.008
Perfluorooctanoic acid PFOA	μg/L	<0.001
6:2 FTS	μg/L	<0.001
8:2 FTS	µg/L	<0.002
Surrogate <sup>13</sup> C <sub>8</sub> PFOS	%	102
Surrogate 13 C2 PFOA	%	88
Extracted ISTD 13 C <sub>3</sub> PFBS	%	73
Extracted ISTD 18 O2 PFHxS	%	96
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOS	%	88
Extracted ISTD 13 C4 PFOA	%	106
Extracted ISTD <sup>13</sup> C₂ 6:2FTS	%	112
Extracted ISTD <sup>13</sup> C₂ 8:2FTS	%	116
Total Positive PFHxS & PFOS	μg/L	0.015
Total Positive PFOA & PFOS	μg/L	0.008
Total Positive PFAS	μg/L	0.017

Envirolab Reference: 369941

Revision No:

Method ID	Methodology Summary
Org-029	Soil samples are extracted with basified Methanol. Waters and soil extracts are directly injected and/or concentrated/extracted using SPE. TCLPs/ASLP leachates are centrifuged, the supernatant is then analysed (including amendment with solvent) - as per the option in AS4439.3.
	Analysis is undertaken with LC-MS/MS.
	PFAS results include the sum of branched and linear isomers where applicable.
	Please note that PFAS results are corrected for Extracted Internal Standards (QSM 5.4 Table B-15 terminology), which are mass labelled analytes added prior to sample preparation to assess matrix effects and verify processing of the sample. PFAS analytes without a commercially available mass labelled analogue are corrected vs a closely eluting mass labelled PFAS compound. Surrogates are also reported, in this context they are mass labelled PFAS compounds added prior to extraction but are used as monitoring compounds only (not used for result correction). Envicarb (or similar) is used discretionally to remove interfering matrix components.
	Please contact the laboratory if estimates of Measurement Uncertainty are required as per WA DER.

Envirolab Reference: 369941 Revision No R00

QUALITY CONTR	OL PFAS I	n Water LC	W LEVEL Short			Du	plicate		Spike Rec	overy %
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date prepared	-			08/01/2025					07/01/2025	
Date analysed	-			08/01/2025	ONT I		100	l mag	07/01/2025	
Perfluorobutanesulfonic acid	μg/L	0.001	Org-029	<0.001			100		102	
Perfluorohexanesulfonic acid - PFHxS	μg/L	0.001	Org-029	<0.001					101	
Perfluorooctanesulfonic acid PFOS	μg/L	0,001	Org-029	<0.001					100	
Perfluorooctanoic acid PFOA	μg/L	0.001	Org-029	<0.001				100	98	
6:2 FTS	μg/L	0.001	Org-029	<0.001					98	
B:2 FTS	μg/L	0.002	Org-029	<0.002			pin	1115	100	
Surrogate <sup>13</sup> C <sub>8</sub> PFOS	%		Org-029	98			207	No.	98	
Surrogate <sup>13</sup> C₂ PFOA	%		Org-029	91	but !		_0776	193	93	
Extracted ISTD <sup>13</sup> C <sub>3</sub> PFBS	%		Org-029	70			10.5	nn.	64	
Extracted ISTD 18 O <sub>2</sub> PFHxS	%		Org-029	80	a)E)		16-2	l PS	77	
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOS	%		Org-029	72			m.		71	
Extracted ISTD 13 C <sub>4</sub> PFOA	%		Org-029	89	HYT.		milit	deta	84	
Extracted ISTD 13 C <sub>2</sub> 6:2FTS	%		Org-029	95	рит		PIE	NG.	83	
Extracted ISTD <sup>13</sup> C <sub>2</sub> 8:2FTS	%		Org-029	93	beri		Ост	(NIT)	83	

Envirolab Reference: 369941 Revision No: R00

Result Definiti	ons
NT	Not tested
NA	Test not required
INS	Insufficient sample for this test
PQL	Practical Quantitation Limit
<	Less than
>	Greater than
RPD	Relative Percent Difference
LCS	Laboratory Control Sample
NS	Not specified
NEPM	National Environmental Protection Measure
NR	Not Reported

Envirolab Reference: 369941 Revision No: R00

Quality Contro	ol Definitions
Blank	This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.
Duplicate	This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.
Matrix Spike	A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.
LCS (Laboratory Control Sample)	This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.
Surrogate Spike	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.

Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011.

The recommended maximums for analytes in urine are taken from "2018 TLVs and BEIs", as published by ACGIH (where available). Limit provided for Nickel is a precautionary guideline as per Position Paper prepared by AIOH Exposure Standards Committee, 2016.

Guideline limits for Rinse Water Quality reported as per analytical requirements and specifications of AS 4187, Amdt 2 2019, Table 7.2

### Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was insufficient in order to satisfy laboratory QA/QC protocols.

When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Where matrix spike recoveries fall below the lower limit of the acceptance criteria (e.g. for non-labile or standard Organics <60%), positive result(s) in the parent sample will subsequently have a higher than typical estimated uncertainty (MU estimates supplied on request) and in these circumstances the sample result is likely biased significantly low.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

Samples for Microbiological analysis (not Amoeba forms) received outside of the 2-8°C temperature range do not meet the ideal cooling conditions as stated in AS2031-2012.

Envirolab Reference 369941

Revision No.